

## Lab 8: Soil respiration using portable chamber

Respiration is the 2<sup>nd</sup> most important carbon flux in ecosystems following GPP (gross primary productivity); and soil respiration makes up 40 to 80% of ecosystem respiration. The objective of this lab is to better understand how environmental constraints (temperature and moisture) and management, effect soil respiration.

### 11.1 Symbols, constants and units

Symbol	Name	units	notes
$T_s$	Soil temperature	°C (or K)	measured
$T_{ref}$	Reference soil temperature	°C (or K)	measured
$V$	Volume of chamber	0.00065363 m <sup>3</sup>	constant
$A$	Area of ground covered by chamber	0.007854 m <sup>2</sup>	constant
$Q_{10}$	Ecosystem respiration	Dimensionless	Equation 5
$\rho$	Molar density of air	40 mol m <sup>-3</sup>	constant
$GEP$	Gross ecosystem photosynthesis	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Equation 1
$NEP$	Net ecosystem productivity	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Equation 1
$NEE$	Net ecosystem exchange	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Equation 2
$R_e$	Ecosystem respiration	$\mu\text{mol m}^{-2} \text{s}^{-1}$	
$R_s$	Soil respiration	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Equation 4
$R_h$	Heterotrophic respiration	$\mu\text{mol m}^{-2} \text{s}^{-1}$	
$R_{rhiz}$	Rhizosphere respiration	$\mu\text{mol m}^{-2} \text{s}^{-1}$	
$F$	Soil CO <sub>2</sub> efflux	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Equation 6

## 11.2 Theory

Carbon sequestration at the ecosystem scale, also known as Net Ecosystem Productivity (NEP) is the difference between photosynthesis and respiration and can be written as:

$$\text{NEP} = \text{GEP} - R_e \quad [1]$$

Thus when NEP is positive ( $\text{GEP} > R_e$ ) the ecosystem is gaining  $\text{CO}_2$ . Alternatively, the net exchange of  $\text{CO}_2$  between terrestrial ecosystems and the atmosphere, called net ecosystem exchange (NEE), is the difference between ecosystem respiration  $R_e$  and gross ecosystem photosynthesis (GEP):

$$\text{NEE} = R_e - \text{GEP} \quad [2]$$

Thus when NEE is positive ( $R_e > \text{GEP}$ ) the atmosphere is gaining  $\text{CO}_2$  (i.e. ecosystem is losing  $\text{CO}_2$ ). NEP is then the net uptake of  $\text{CO}_2$  by the ecosystem from the atmosphere (a term used by biologists), while NEE (the term used by biometeorologists) is:

$$\text{NEE} = - \text{NEP} \quad [3]$$

Soil respiration ( $R_s$ ) can account for as much as 80% of ecosystem respiration, and results in the efflux of  $\text{CO}_2$  from the soil.  $R_s$  is composed of the  $\text{CO}_2$  produced by microbial decomposition of soil organic matter (SOM) – heterotrophic respiration ( $R_h$ ), and by rhizosphere respiration ( $R_{rhiz}$ ) associated with root respiration + mycorrhizal and microbes in the rhizosphere fed by root exudates.

$$R_s = R_h + R_{rhiz} \quad [4]$$

Soil temperature ( $T_s$ ) followed by soil moisture ( $\Theta$ ) are the most important environmental factors affecting  $R_e$ .  $R_e$  increases exponentially with  $T_s$ , often expressed as:

$$R = R_{ref} Q_{10}^{\left(\frac{T_s - T_{ref}}{10}\right)} \quad [5]$$

where:  $R_{ref}$  is  $R_e$  at a reference temperature  $T_{ref}$ , usually  $10^\circ\text{C}$ ; and  $Q_{10}$  is the relative increase in  $R_e$  for a 10 degree increase in  $T$ .  $Q_{10}$  is about 2 in many biological processes,

but may be as high as 3 for  $R_e$  and  $R_s$ . This sensitivity is particularly important in predicting the impacts of global warming on decomposition rates, but is dependent on substrate quantity and quality, soil temperature and soil moisture.

A widely used method for measuring  $R_s$  is the closed or static chamber method. In this method the rate of  $\text{CO}_2$  concentration in a chamber placed on the soil surface is used to determine the soil  $\text{CO}_2$  efflux ( $F$ ). This is a good measure of  $F$  as little respired  $\text{CO}_2$  accumulates in the soil. Thus  $F$  can be calculated using:

$$F = \frac{\rho V}{A} \frac{dC}{dt} \quad [6]$$

where:  $\rho$  is the molar density of air ( $40 \text{ mol m}^{-3}$ ),  $V$  is the volume of the chamber ( $\text{m}^3$ ),  $A$  is the area of ground covered by the chamber ( $\text{m}^2$ ),  $d\text{CO}_2/dt$  is the rate of increase in  $\text{CO}_2$  concentration, and  $F$  is in units of  $\mu\text{mol m}^{-2} \text{ s}^{-1}$ . If the chamber is kept on the soil < 2 minutes,  $d\text{CO}_2/dt$  is almost constant (linear); that is the increase in  $\text{CO}_2$  concentration does not significantly suppress  $\text{CO}_2$  efflux. As the  $\text{CO}_2$  concentration changes during the measurement, this method is described as non-steady state.

Recall that  $\text{NEE} = R_e - \text{GEP}$ . So if there is vegetation in your chamber you are measuring  $\text{NEE}$ . If there is not vegetation  $\text{GEP} \rightarrow 0$  and  $R_e = R_s$ .

### 11.3 Objectives

To determine the sensitivity of  $R_s$ , by comparing the magnitude of  $F$  from bare agricultural soil, (both fertilized and unfertilized), and a soil under a cover crop (both fertilized and unfertilized). Four containers of soils have been collected from the Delta region, which are Gleysolic soils with silty clay loam texture. Two containers have been seeded with a typical winter cover crop in the region and two have been left bare. One bare soil and one cover cropped soil have been fertilized with urea fertilizer (46-0-0). All soils have been watered equally to obtain a soil volumetric water content close to field capacity (25-35%).

## 11.4 Laboratory Procedure

1. Allow the instructor/assistant to bring up the soil greenhouse gas software.
2. Plug in your Vaisala GMP252 Carbon dioxide probe into an electrical socket.
3. Wait 1-2 minutes for the sensor to start measuring, and for the screen to appear on the projector screen.
4. Ensure that the Vaisala GMP252 Carbon dioxide probe's chamber opening is flat on the table. Allow the probe a sufficient amount of time to equilibrate to atmospheric conditions – this can be recognized by a flat reading approximately 400-500 ppm for 2-3 minutes. This typically can take 4-5 minutes from startup of the probe.
5. At the top of the measurement sheet record the soil treatment you are working on.
6. Using the display screen
  - a. Using the rubber end of the pencil you can gently tap on the display to perform certain functions.
    - i. **Resize the graph** – tapping in the top left-hand corner



- ii. **Refresh the graph** – tapping in the top right-hand corner



- iii. **Obtain an estimate flux** – tapping at the start of the increase in the CO<sub>2</sub> graph will allow you to select a point. It will then generate a line and slope automatically using a 60 second time interval. This slope is an approximate value for the flux of CO<sub>2</sub>.



7. Once the probe has equilibrated, record the ppm reading on your datasheet (this is your initial value at time 0). Set up a timer on your smartphone or a stopwatch.
8. Gently pick the sensor up and place it onto the soil gas collar that has already been inserted into the soil. Be sure to go slow, so as to not create a significant pressure differential inside the chamber. Simultaneously start your timer.
9. Every 30 seconds, record the concentration of carbon dioxide (ppm) displayed on the CO<sub>2</sub> sensor screen for 180 seconds.

10. After 180 seconds, remove the chamber from the collar and place the chamber on the counter with the open side facing out and away from people, allow it to reach equilibrium again (3-4 minutes). Your reading value should approach your initial value.
11. Using the eraser end of the pencil, gently tap on the screen at the start of the increase in CO<sub>2</sub> concentration where there is a relatively straight line. This will give you an approximate CO<sub>2</sub> flux. You can tap on the top left hand of the screen to resize the curve if needed, and refresh the screen by tapping on the top right hand side. Record this readout flux value from the sensor. You will need to get 3 relatively consistent readings for each soil/vegetation/fertilizer combination (i.e. at each station).
12. Insert one thermometer approximately 5 cm into the soil and record the temperature after 3 minutes.
13. Leave one thermometer on the counter, and record the temperature of the air at 3 minutes.
14. Use the TDR to measure soil volumetric water content at 3 points around the soil gas collar. Ensure that the measurement mode is set to the correct soil type for your experiment.
15. Flip the sensor flat onto the table for 2-3 minutes until it reaches a constant measurement. Note: the equilibrated concentration of CO<sub>2</sub> may increase slightly in the room due to the amount of people in the room respiring. Use a fan to assist the small fan in the unit to re-circulate air into the headspace of the chamber if necessary.
16. Repeat steps 7-15 on each soil treatment a minimum of 3 times in order to obtain an average gas flux measurement. Note if there is inconsistency in your flux values determined from the readout screen, you will need to repeat measurements > 3 times.
17. After obtaining an average CO<sub>2</sub> flux for your soil (bare/vegetated, +/- fertilizer) all groups will rotate to the next station and repeat measurements for that soil.

## 11.5 Calculations

1. Plot the change in ppm versus time (s) for each of your 4 soils (and replicates), and calculate the slope  $\Delta\text{CO}_2/\Delta t$  ( $\text{ppm CO}_2 \text{ s}^{-1}$ )

For each soil, convert the measured  $\text{CO}_2$  increase ( $\text{ppm CO}_2 \text{ s}^{-1}$ ), to flux density,  $F$  ( $\mu\text{mol m}^{-2} \text{ s}^{-1}$ ) using equation 6. Obtain values for  $p$ ,  $V$  and  $A$  from section 2.1 symbols, values and units.

Complete data sheet (and attached to your assignment)

## 11.6 References

Davidson E.A. Janssens I.A. 2006. Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature* 440(9): 165-173.

Jassal R. Black A. Novak M. Morgenstern K. Nesic Z. Gaumont-Guay D. 2005. Relationship between soil  $\text{CO}_2$  concentrations and forest-floor  $\text{CO}_2$  effluxes. *Agricultural and Forest Meteorology* 130:176-192.

Jassal R. Black A. Novak M. Gaumont-Guay D. Nesic Z. 2008. Effect of soil water stress on soil respiration and its temperature sensitivity in an 18-year-old temperate Douglas-fir stand. *Global Change Biology* 14(6): 1305-1318.

### Soil Respiration Lab

Reading Number	Covered Soil		Bare Soil		Covered Soil With Fertilizer		Bare Soil With Fertilizer	
	time (s)	CO2 (ppm)	time (s)	CO2 (ppm)	time (s)	CO2 (ppm)	time (s)	CO2 (ppm)
1	0		0		0		0	
	30		30		30		30	
	60		60		60		60	
	90		90		90		90	
	120		120		120		120	
	150		150		150		150	
	180		180		180		180	
	Slope on display:		Slope on display:		Slope on display:		Slope on display:	
	Slope calculation:		Slope calculation:		Slope calculation:		Slope calculation:	
2	0		0		0		0	
	30		30		30		30	
	60		60		60		60	
	90		90		90		90	
	120		120		120		120	
	150		150		150		150	
	180		180		180		180	
	Slope on display:		Slope on display:		Slope on display:		Slope on display:	
	Slope calculation:		Slope calculation:		Slope calculation:		Slope calculation:	
3	0		0		0		0	
	30		30		30		30	
	60		60		60		60	
	90		90		90		90	
	120		120		120		120	
	150		150		150		150	
	180		180		180		180	
	Slope on display:		Slope on display:		Slope on display:		Slope on display:	
	Slope calculation:		Slope calculation:		Slope calculation:		Slope calculation:	



