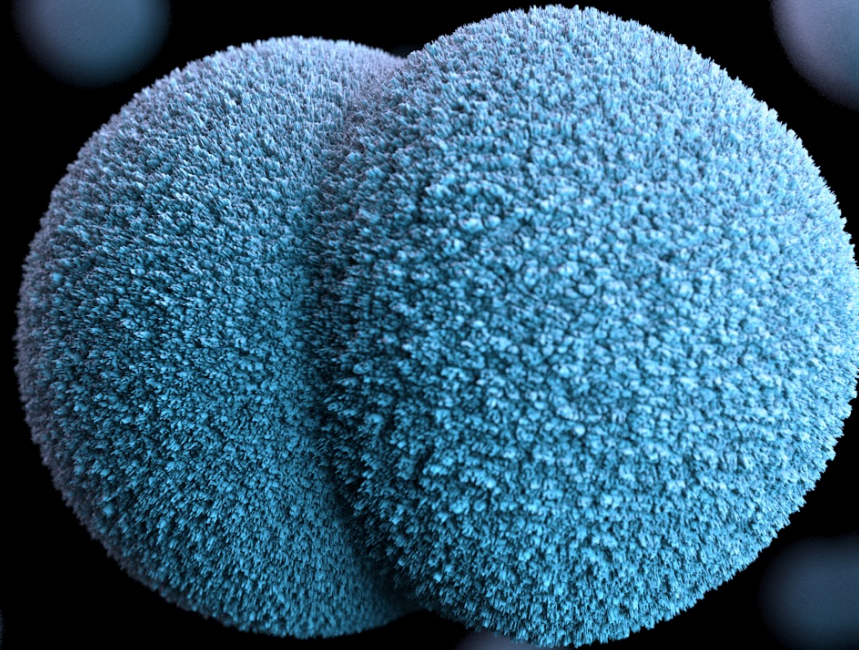


IMAGE SOURCE: CDC¹

3D, COMPUTER-GENERATED IMAGE OF *NEISSERIA MENINGITIDIS*
DIPLOCOCCI BASED ON SEM IMAGERY



CASE 1: A STIFF NECK

MICROBIOLOGY LABORATORY SUMMARY

CLAIRE SIE

PATH417 2021W2

CASE OVERVIEW



“18-year-old Mary has just moved into the dormitory at her university. One day, her roommate finds her lying in bed under her sheets. She is complaining of fever, chills, bad headache, and a stiff neck. She is staying under the covers because the light is hurting her eyes. Her roommate calls 911 and an ambulance takes Mary to the local hospital.

The emergency room physician asks Mary about her recent vaccinations, and she reports that she has not had any since she was in elementary school. The physician documents a fever of 39.2C and low blood pressure. He sends blood and cerebral spinal fluid to the Microbiology Laboratory. She is started immediately on intravenous antibiotics. Mary’s blood and cerebral spinal fluid grow *Neisseria meningitidis* and she is diagnosed with meningococcal meningitis.”

WHAT IS MENINGITIS?



INFLAMMATION OF THE MENINGES

Meninges – protective membrane surrounding brain and spinal cord



CAN CAUSE DAMAGE TO THE CNS

Increased lymphocyte invasion, neuron damage, and intracranial swelling



VARIETY OF PATHOGENIC CAUSES

Bacterial, fungal, viral – treatments vary

Symptoms:



FEVER AND CHILLS

Elevated body temperature



PAIN

Headache and stiff neck



PHOTOPHOBIA

Eyes become sensitive to light

MAIN CAUSATIVE AGENT: *NEISSERIA MENINGITIDIS*

- ❖ Gram-negative, encapsulated, human pathogen
- ❖ Leading cause of bacterial meningitis worldwide¹
- ❖ Commensal organism in ~10% of population²
- ❖ 13 serotypes based on variation in porin protein (6 disease-causing)³
- ❖ Invades and replicates in cerebral spinal fluid (CSF)
- ❖ Capsule-based, conjugate vaccines available and effective

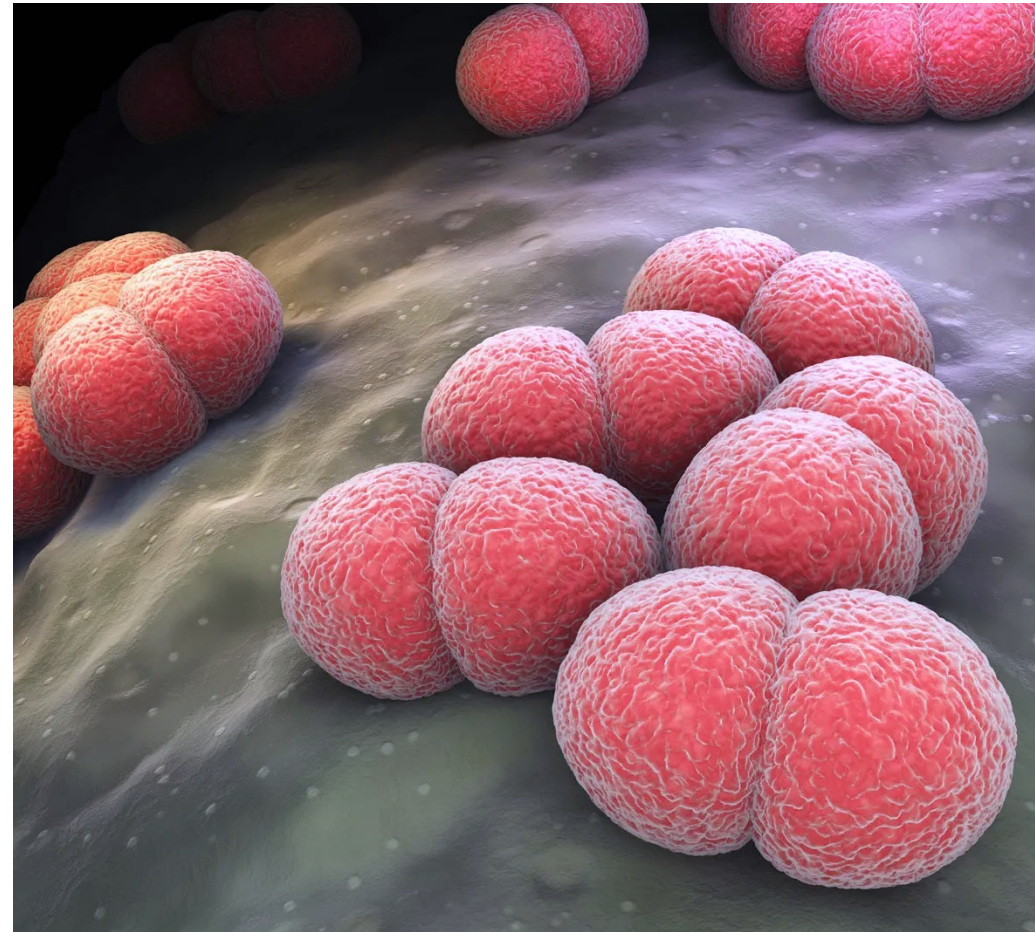
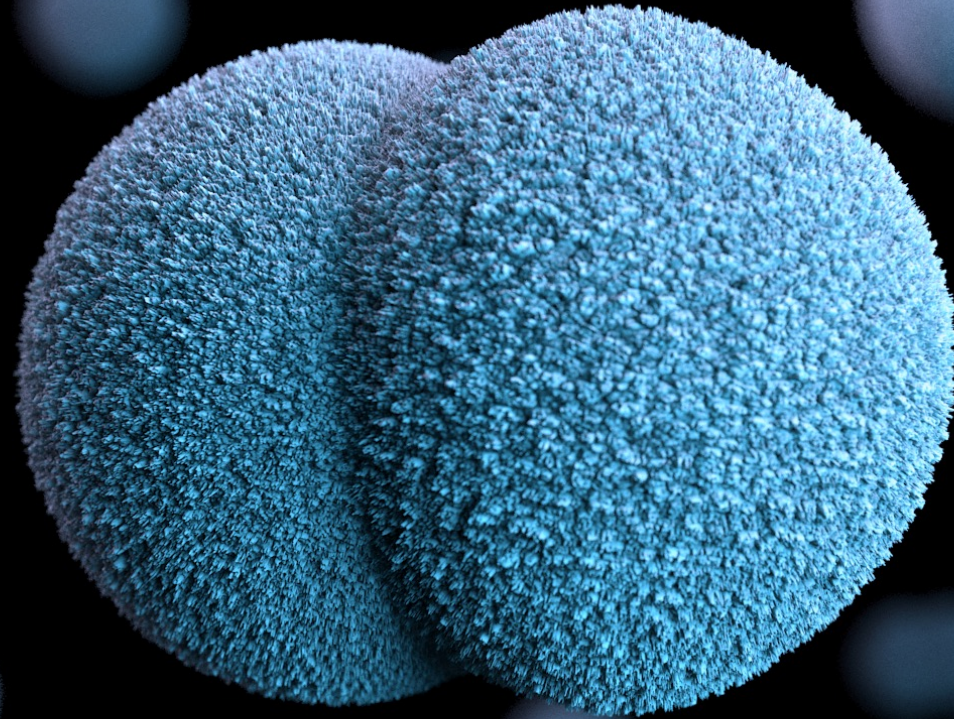


IMAGE SOURCE: CDC¹

3D, COMPUTER-GENERATED IMAGE OF *NEISSERIA MENINGITIDIS*
DIPLOCOCCI BASED ON SEM IMAGERY



COMMON PATHOGENS

Other than the stated bacterial cause, what are the most common bacterial pathogens associated with this type of infectious scenario?

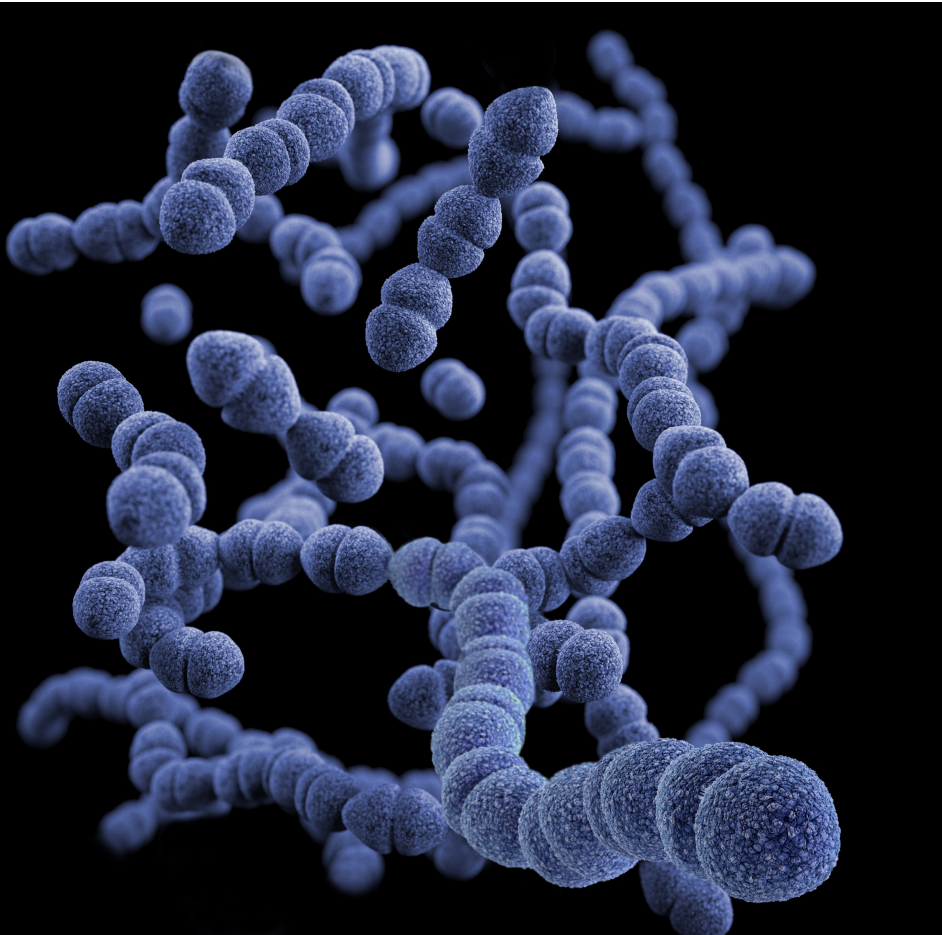
POTENTIAL CAUSATIVE AGENTS

1. *Neisseria meningitidis*
2. *Streptococcus pneumoniae*
3. *Haemophilus influenzae*
4. Group B streptococcus
5. *Escherichia coli*
6. *Listeria monocytogenes*

Responsible for majority of meningitis cases in the US (all age groups)³

More common causative agent of bacterial meningitis in infants younger than 30 days⁴

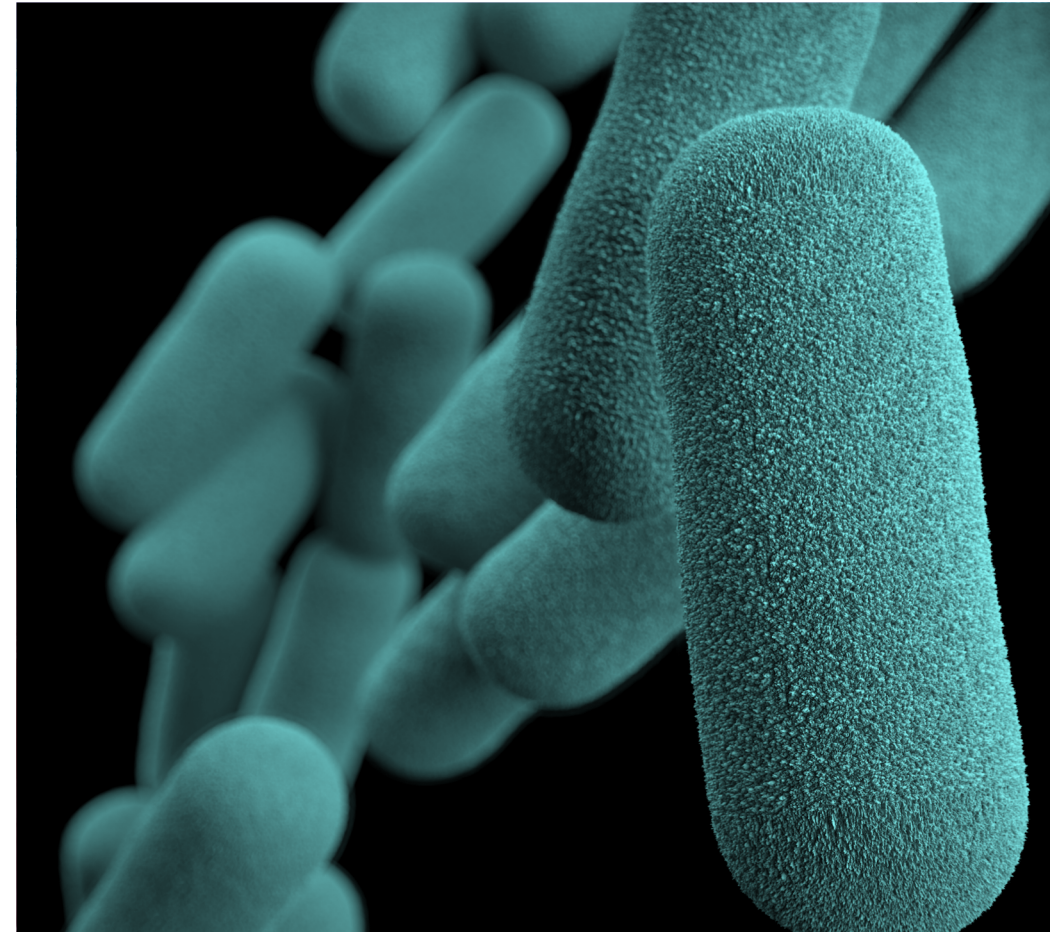
STREPTOCOCCUS PNEUMONIAE



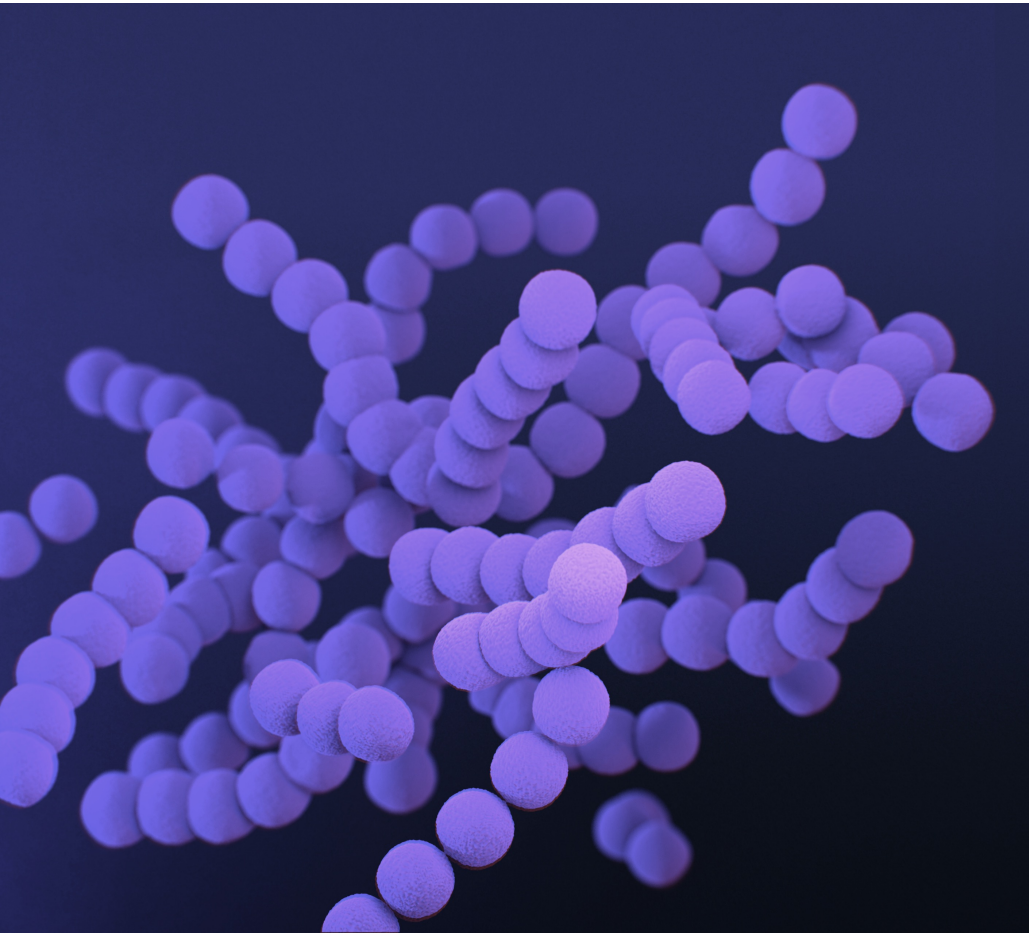
- ❖ Gram-positive, lancet-shaped pathogen
- ❖ Causative agent in 58% of meningitis cases in US⁵
- ❖ Can cross blood-brain barrier to infect central nervous system (CNS)⁶
- ❖ Asymptomatic colonization of upper respiratory tract in 5-10% of adults without children⁷
- ❖ Transmission through respiratory droplets
- ❖ Increasing rates of penicillin resistance⁸
- ❖ Preventative vaccine available⁹

HAEMOPHILUS INFLUENZAE

- ❖ Gram-negative, encapsulated pathogen
- ❖ Subgroup division based on capsule type, *H. influenza* Type B (Hib) dominant¹⁰
- ❖ Particular risk of infection in patients <5 and >65, immunocompromised individuals¹⁰
- ❖ Increasing rates of ampicillin resistance¹⁰
- ❖ Can asymptotically colonize nasopharynx¹¹
- ❖ Transmission through respiratory droplets



GROUP B STREPTOCOCCUS



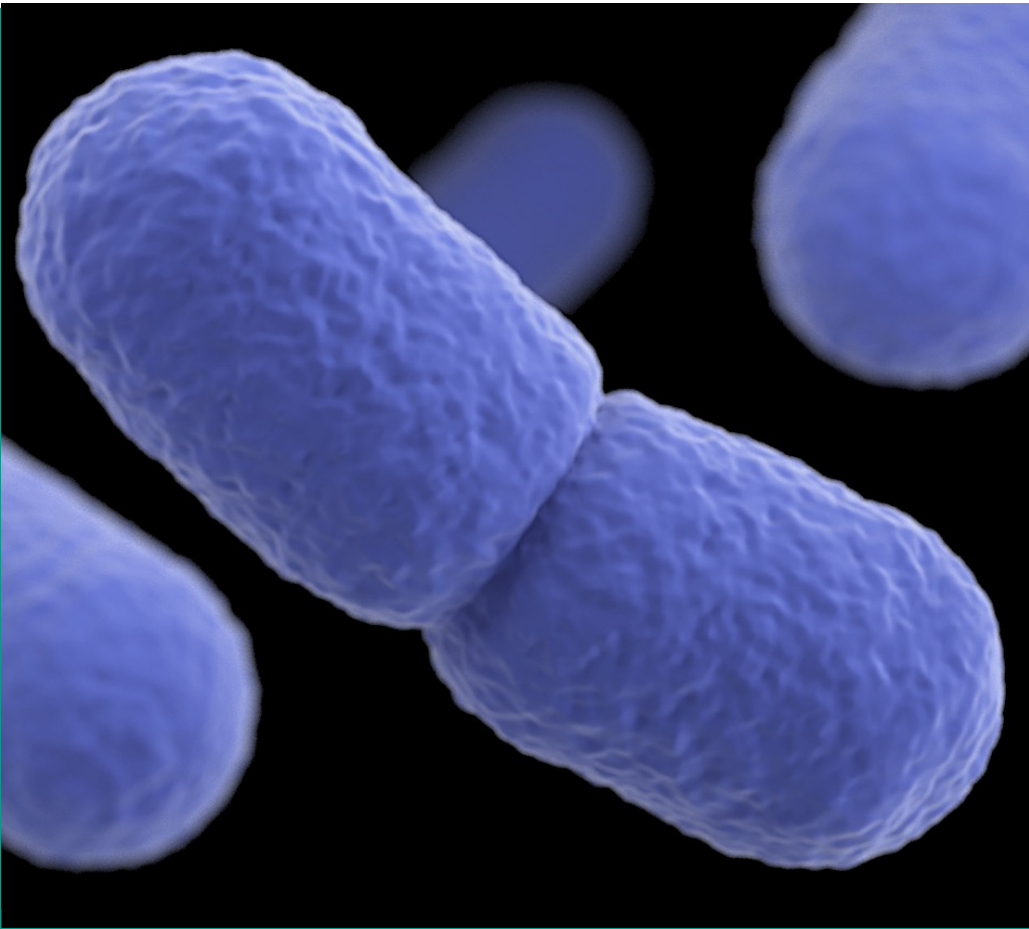
- ❖ Gram-positive cocci, forms pairs or chains
- ❖ Commensal organism in reproductive and GI tract in 1/3 of women¹²
- ❖ Transmission to newborns during childbirth¹³
- ❖ Contact-based transmission is unlikely¹³ – therefore Mary is unlikely to be infected with it

ESCHERICHIA COLI

- ❖ Gram-negative organism
- ❖ Establishes infection in infants due to immature immune system
- ❖ Meningitis-causing strains possess K1 polysialic capsule – essential for crossing BBB¹⁴
- ❖ Typically harmless and commensal in intestines¹⁵
- ❖ Transmission to newborn occurs during vaginal delivery¹⁶
- ❖ Unlikely to be causative agent in Mary's case (*E. coli* meningitis rare in adults¹⁷)



LISTERIA MONOCYTOGENES



- ❖ Gram-positive, intracellular pathogen
- ❖ Transmission by ingestion of contaminated food, or during vaginal delivery¹⁸
- ❖ Environmental factors – common cause of bacterial meningitis in sub-Saharan Africa¹⁹
- ❖ Listeriosis – additional symptoms include convulsions and loss of balance¹⁷
- ❖ Unlikely to be causative agent in Mary's case due to her lack of additional symptoms

IMAGE SOURCE: PEXELS⁹ 2021. IMAGE FREE FOR USE.

BLOOD SAMPLES IN COLLECTION TUBES



SAMPLING

What samples are taken for laboratory testing and how important is the Microbiology Laboratory in the diagnosis of this particular infectious disease?

IMPORTANCE OF THE MICROBIOLOGY LABORATORY

Clinicians will collect samples for a three-pronged approach to microbiological testing:

- Blood culture
- Cerebrospinal fluid (CSF) Gram stain
- CSF culture

Effectively and correctly diagnoses 90% of cases with Mary's symptoms (fever, headache, stiff neck)²⁰

Important to identify the causative agent of Mary's symptoms as different pathogens are sensitive/resistant to different antibiotics

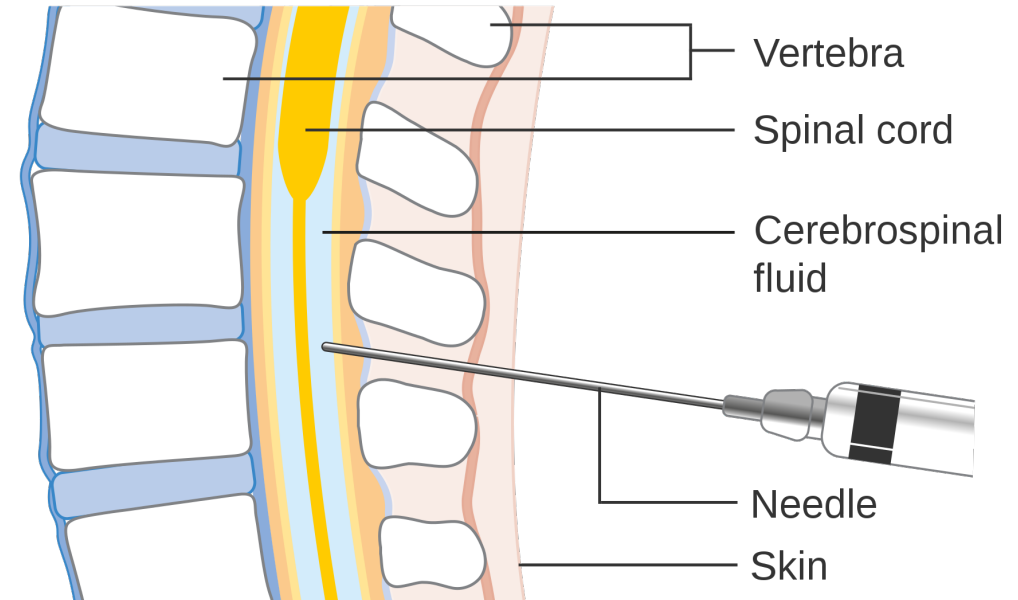
Treatment with broad-spectrum antibiotics can lead to failure to clear pathogen and development of antibiotic resistance!

CEREBROSPINAL FLUID (CSF)

A CLEAR AND COLOURLESS FLUID WHICH CUSHIONS AND PROVIDES NUTRIENTS TO THE CENTRAL NERVOUS SYSTEM (CNS)

Collection Process:

- Sample collected before or shortly after administration of [antibiotics](#)
- Obtained via [lumbar puncture](#)
- Needle inserted between lumbar vertebrae (image shown) while patient is sitting or on their side; sterility important – contamination can interfere with testing results
- Sample taken to Microbiology Laboratory for analysis [within an hour](#) of collection²¹
 - If not possible, inoculate into Trans-Isolate (T-I) medium and incubation at 5% CO₂, 37C until ready for transport²¹



Examine CSF for:

- Cell count
- Glucose level
- Gram stain
- Cultures
- Polymerase Chain Reaction (PCR)

BLOOD

Collection Process:

- ❑ 1-3 ml collected before or shortly after administration of [antibiotics](#)²²
- ❑ Sample should be [inoculated within one minute](#) of collection and taken to Microbiology Laboratory immediately²¹

Used for:

- ❑ Complete blood count (CBC)
- ❑ Coagulation studies
- ❑ Electrolyte levels
- ❑ Probe for inflammatory markers



NOTES ON SAMPLE COLLECTION

- ❖ If samples were collected after antibiotic administration, or if no bacteria observed via Gram stain:
 - ❖ Perform [Polymerase Chain Reaction test](#) (PCR) -
 - ❖ PCR can confirm:
 - ❖ Meningitis
 - ❖ Causative agent - e.g. *N. meningitidis*, *S. pneumoniae*, *H. influenzae*²³
- ❖ Important! Follow biosafety guidelines to reduce risk of being infected with unknown pathogen while collecting CSF and blood samples²¹
 - ❖ Wash hands, wear gloves, sterile technique, safe sample transport, safe sharps disposal, immediate and complete reporting of any injuries or contamination

IMAGE SOURCE: PIXABAY¹² 2017. IMAGE FREE FOR COMMERCIAL USE. NO ATTRIBUTION REQUIRED.

A CLOSE-UP IMAGE OF A MICROSCOPE

LABORATORY TESTING

Explain the tests that will be performed on the samples in order to detect any of the potential bacterial pathogens causing this disease.

TESTS PERFORMED ON SAMPLES



CULTURE

Growth on Blood or Chocolate agar plates, macroscopic and microscopic morphology



BIOCHEMICAL TESTING

Catalase, Oxidase, Carbohydrate utilization, Growth requirements



CSF CYTOLOGICAL ANALYSIS

Glucose level, CSF protein concentration, lymphocyte infiltration



AGGLUTINATION

Identify pathogen using pathogen-specific antisera or antibody-coated latex particles



GRAM STAIN

Identify pathogen as Gram-positive or Gram-negative



POLYMERASE CHAIN REACTION

Identify pathogen based on presence of organism-specific DNA sequences

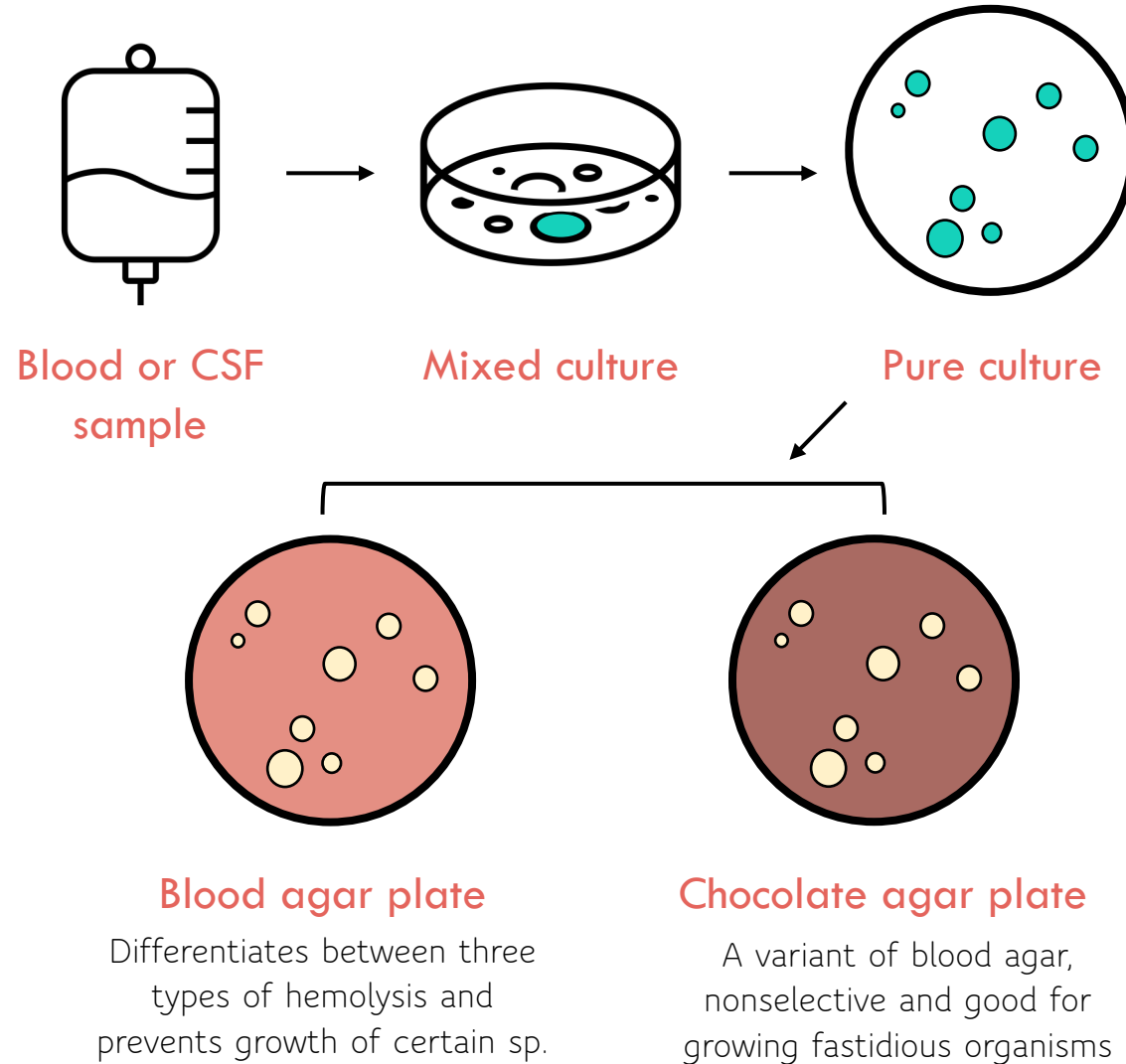
CULTURE

- ❖ Isolate a **pure culture**²⁴ from sample by:
 - ❖ Streak from sample to obtain single colonies
 - ❖ Inoculate blood agar plate or chocolate agar plate from single colony²⁵
 - ❖ For use in further testing
- ❖ Observe **macroscopic morphology**
- ❖ Observe growth on **selective/differential media**

Positive blood culture can confirm²⁴:

- ❑ 40% of meningococcal meningitis cases
- ❑ 50-90% of *H. influenzae* cases
- ❑ 75% of pneumococcal meningitis cases

CULTURE



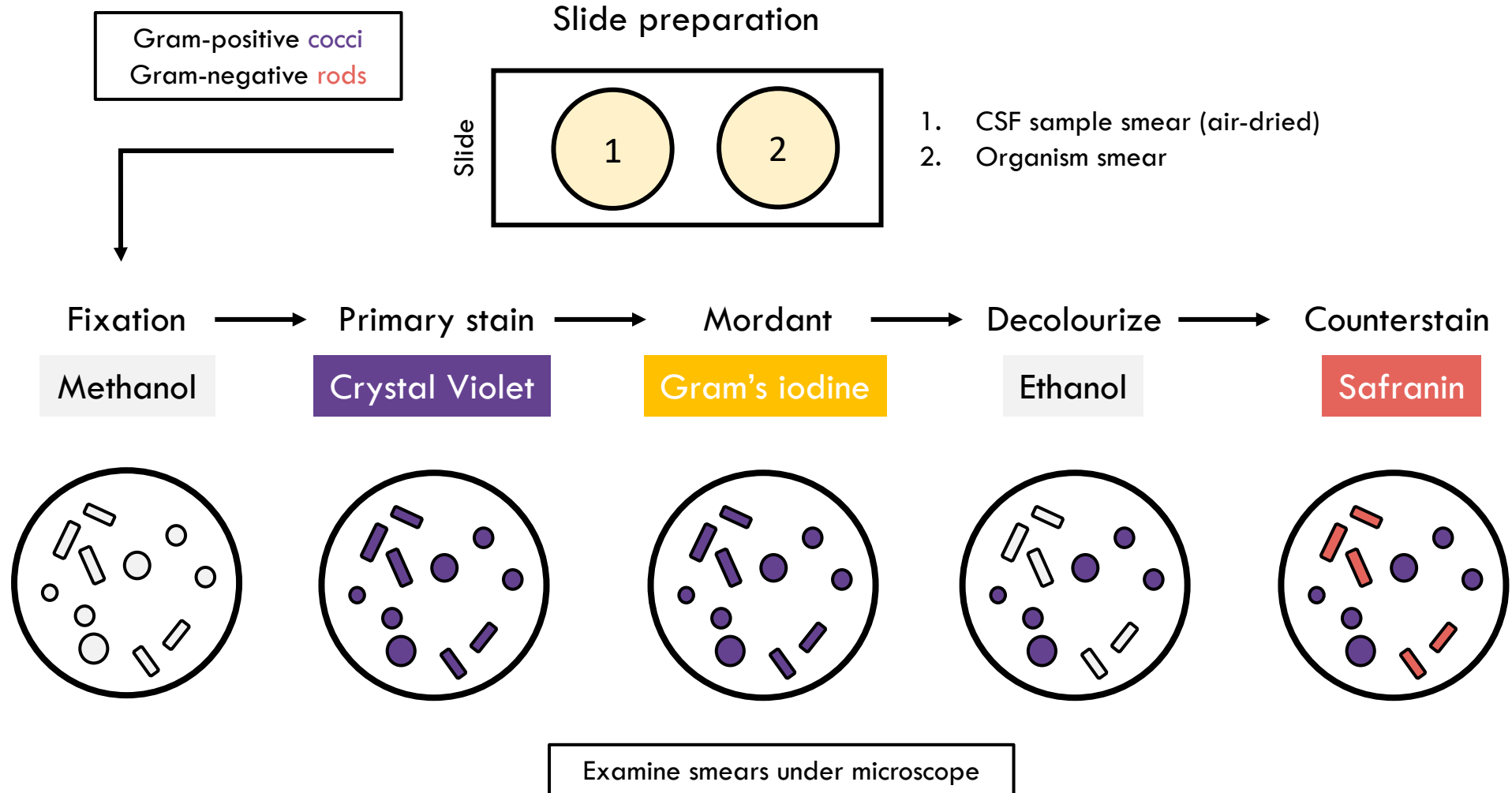
CSF CYTOLOGICAL ANALYSIS

Cerebrospinal fluid property	Healthy child or adult	Bacterial meningitis	Notes
Colour/Turbidity	Clear	Unclear	Presence of WBCs or RBCs ²⁷
Total leukocytes (cells/mm ³)	<6	>1000	Seen in 87% of bacterial meningitis cases ^{20,27}
Neutrophils (%)	0	>85-90%	Indicates inflammatory response
Protein (mg/dL)	20-40	>100-150	Sensitive indicator of CNS disease ²⁷
Glucose level (mg/dL)	40-80	0-<40	0.4 mg/dL 80% indicative of bacterial meningitis ²⁰
Select biomarkers (C-reactive protein, procalcitonin)		Elevated	Elevated levels point to bacterial meningitis ^{20,28}

Perform blood cultures in conjunction with CSF cytological analysis and CSF culture

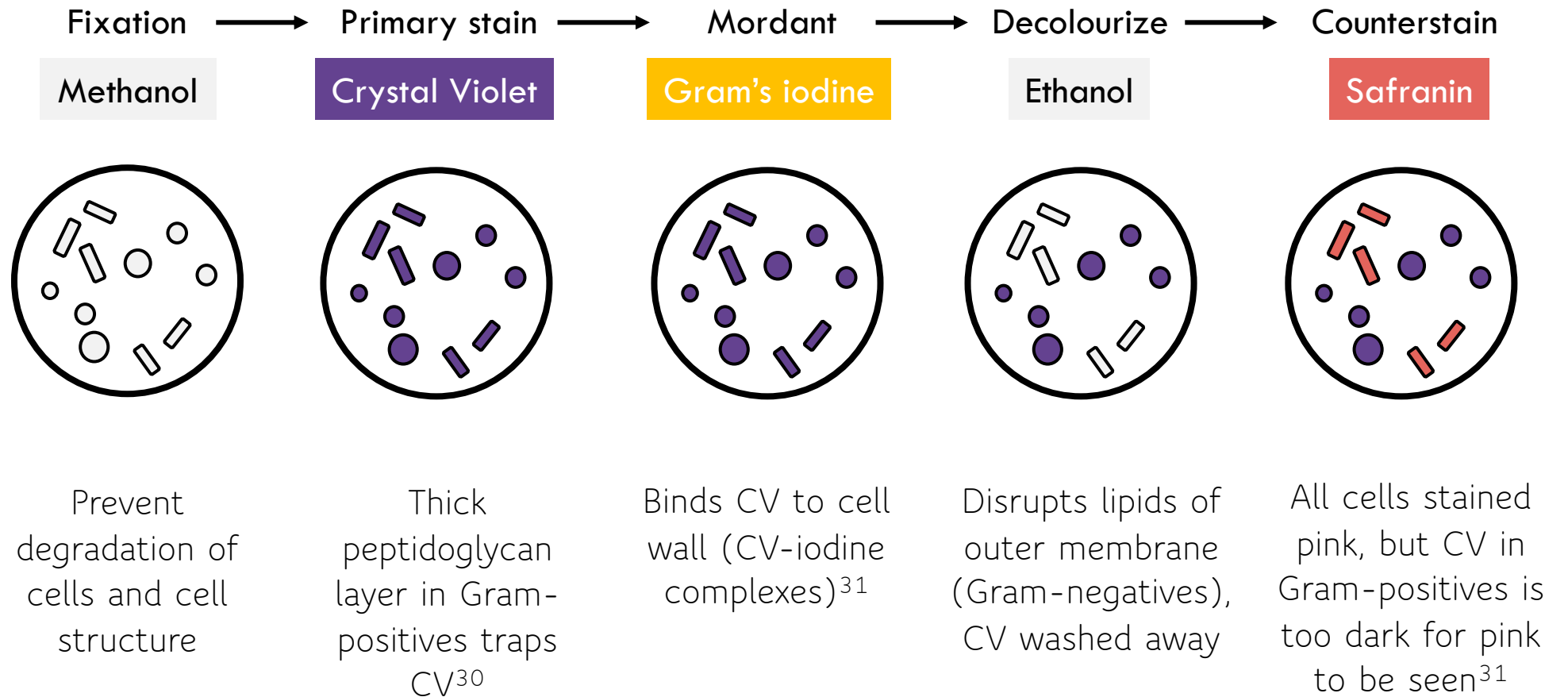
GRAM STAIN PROCEDURE

DIFFERENTIATE BETWEEN ORGANISMS BASED ON BACTERIAL CELL WALL PROPERTIES²⁹



GRAM STAIN

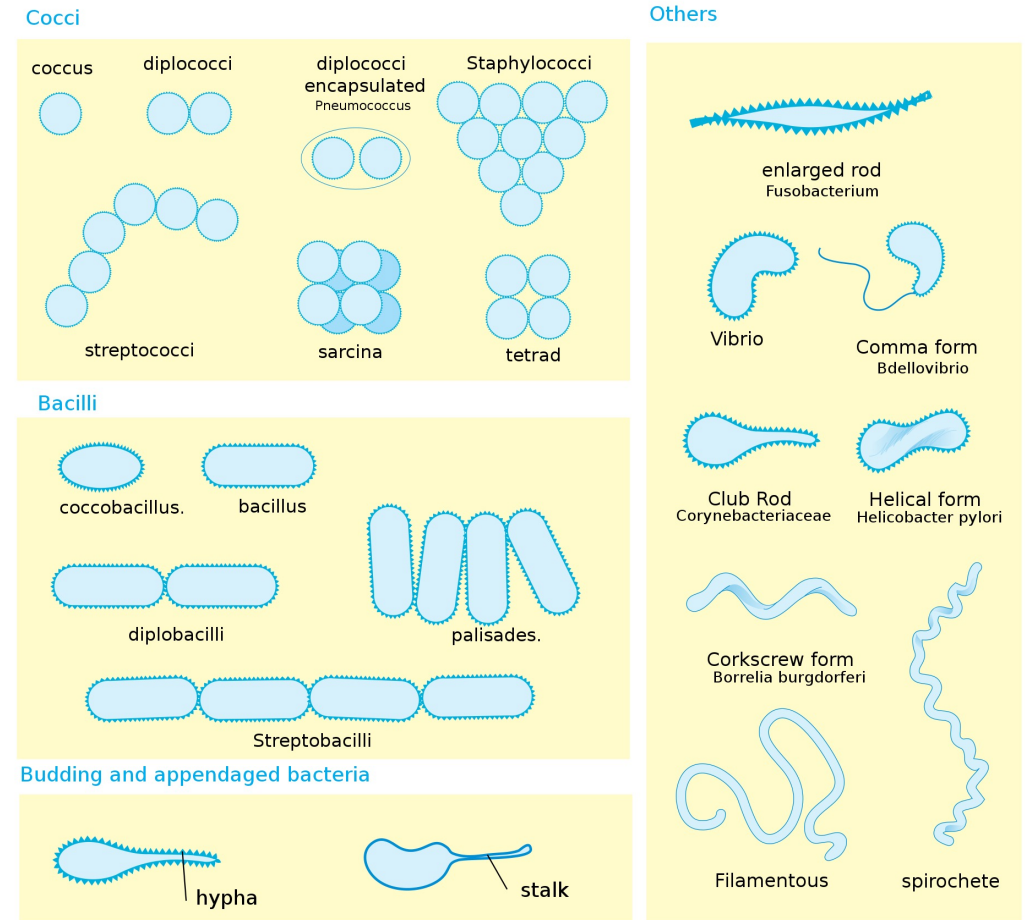
DIFFERENTIATE BETWEEN ORGANISMS BASED ON BACTERIAL CELL WALL PROPERTIES



MICROSCOPIC MORPHOLOGY

CAN BE DONE CONCURRENTLY WHILE OBSERVING RESULTS OF GRAM STAIN

- ❖ Bacterial organisms exhibit **characteristic morphological differences** which allow them to be identified
 - ❖ Cell **shape** (shown right)
 - ❖ Cell **size**
 - ❖ Cell clustering **arrangements**
 - ❖ Additional structures (endospores, capsules)
- ❖ E.g. *N. meningitidis* appears most often as **coffee-bean-shaped diplococci**, whereas *S. pneumoniae* are typically lancet-shaped cocci which have an increased tendency to form **long chains**



BIOCHEMICAL TESTING

Catalase Test

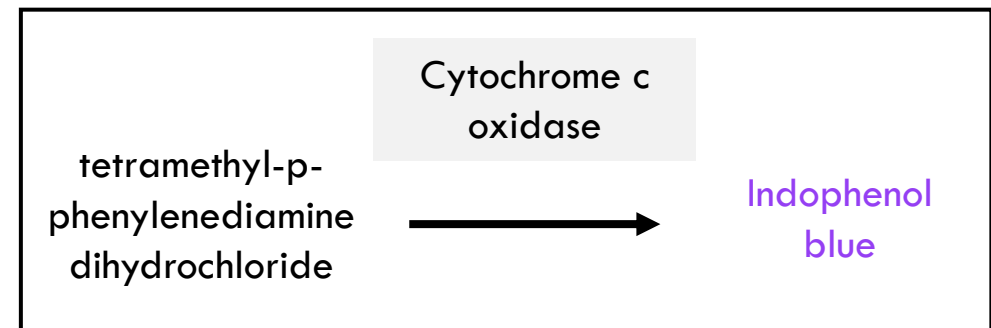
DETECT *S. PNEUMONIAE*

- ❖ **Catalase** decomposes hydrogen peroxide (H_2O_2) to $H_2O + O_2$ = results in **bubbling**³²
- ❖ Discriminate between **Gram-positive cocci**³³
 - ❖ *S. pneumoniae* and *Enterococcus* = negative
 - ❖ Staphylococcus = positive
- ❖ Procedure³¹:
 - ❖ Grow up bacteria on BAP for 24 hours
 - ❖ Using loop, transfer a single colony to glass slide
 - ❖ Add **hydrogen peroxide** and observe for bubbling

Kovac's Oxidase Test

DETECT *N. MENINGITIDIS* AND *H. INFLUENZAE*

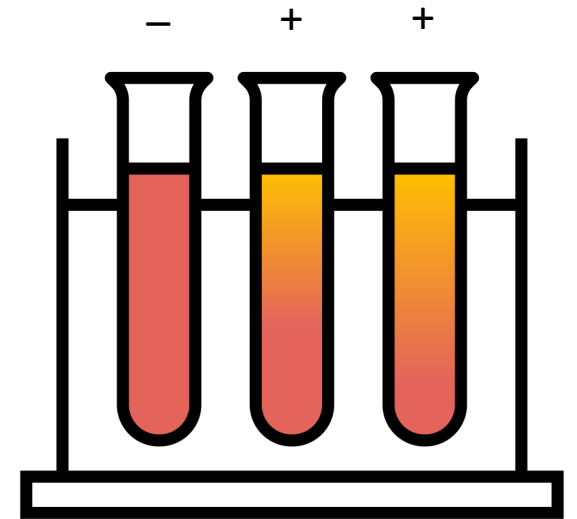
- ❖ Determine presence of **cytochrome c oxidase** in respiratory chain³¹
- ❖ Procedure³¹:
 - ❖ Grow up bacteria on BAP for 24 hours
 - ❖ **Kovac's oxidase reagent** blotted on filter paper, dried
 - ❖ Using loop, transfer a single colony to paper, observe for **colour change**



CARBOHYDRATE UTILIZATION

DETECT *N. MENINGITIDIS*

- ❖ Determine organism's **ability to ferment carbohydrates**
- ❖ *N. meningitidis* only oxidizes glucose and maltose³⁰
- ❖ Add **carbohydrates** to tubes containing cystine trypticase agar (CTA) and **phenol red** (acid indicator)³¹
- ❖ Carbohydrate metabolism/oxidization **produces acid** and sometimes **gas**
- ❖ Procedure³⁰:
 - ❖ Grow up bacteria on BAP for 24 hours
 - ❖ Inoculate CTA-carbohydrate media with a single colony
 - ❖ Incubate at least 72 hours
 - ❖ Observe **colour change** (red → yellow = positive)



OPTOCHIN TEST

DETECT *S. PNEUMONIAE*

- ❖ Optochin = ethylhydrocupreine hydrochloride
- ❖ *S. pneumoniae* strains are sensitive to optochin³²
- ❖ Procedure³²:
 - ❖ Grow up bacteria on BAP for 24 hours
 - ❖ Using a loop, streak out a single colony onto half of a new BAP
 - ❖ Place an **optochin disk** within the streak
 - ❖ Incubate overnight at 35-37C
 - ❖ Measure **zone of inhibition** the following day
 - ❖ >14 mm = organism is sensitive (*S. pneumoniae*)

HEMIN AND NAD GROWTH FACTOR TEST

DETECT *H. INFLUENZAE*

- ❖ *H. influenzae* requires hemin and NAD for growth
- ❖ Procedure³⁴:
 - ❖ Grow up bacteria on BAP for 24 hours
 - ❖ Create a turbid cell suspension (>1.0 McFarland standard) and vortex
 - ❖ Inoculate a trypticase soy agar plate with the cell suspension and allow to dry
 - ❖ Place paper strips/disks containing:
 - ❖ Hemin
 - ❖ NAD
 - ❖ Hemin + NAD
 - ❖ Observe for growth

- ❖ Note: *H. haemolyticus* also requires hemin and NAD for growth³⁴ – test for hemolysis using rabbit or horse blood agar!

AGGLUTINATION TESTING

Agglutination Serogrouping

DETECT *N. MENINGITIDIS* AND *H. INFLUENZAE*

- A.k.a. Slide agglutination serogrouping (SASG)
- Use **serogroup-specific antisera**
- Binding of antisera to bacteria causes **clumps** to form³⁰
- Observe for clumping within 1-2 minutes
- Ratings:
 - Negative: 0, 1+, 2+
 - Positive: 3+, 4+

Latex Agglutination

DETECT *S. PNEUMONIAE*

- Quick detection of bacterial antigens in CSF
- Variable sensitivity
- Bacteria expressing **specific antigens** are mixed with **antibody-coated latex particles**
- Procedure³¹:
 - Centrifuge CSF to remove sediments
 - Heat CSF supernatant to 100C
 - Add 30-50 uL heated CSF to latex reagent
 - Observe for **precipitation** and **clumping** within 5-10 seconds (positive for agglutination)

BILE SOLUBILITY TEST

DETECT *S. PNEUMONIAE*

- ❖ Discriminate *S. pneumoniae* from other streptococci sp.³²
 - ❖ *S. pneumoniae* soluble in bile
 - ❖ Other streptococci insoluble in bile
- ❖ Procedure³²:
 - ❖ Prepare bile solution by dissolving sodium deoxycholate in sterile water
 - ❖ Grow up bacteria on BAP for 24 hours
 - ❖ Transfer an isolated colony into a tube, dilute to 0.5-1.0 McFarland standard turbidity in saline
 - ❖ Add bile solution to cell suspension
 - ❖ Incubate tubes in CO₂
 - ❖ Vortex and observe for turbidity
 - ❖ Positive (bile-soluble): clear

POLYMERASE CHAIN REACTION (PCR) TEST

- ❖ Targets a [pathogen-specific DNA sequence](#)
- ❖ Several technologies exist:
 - ❖ [Traditional PCR](#)
 - ❖ Design primers to specifically bind to and amplify unique DNA sequences³⁵
 - ❖ Visualize amplicons using agarose gel electrophoresis (some concerns with contamination)
 - ❖ [Real-time PCR](#)
 - ❖ Using fluorescently-conjugated oligonucleotide probe which binds unique DNA sequence³⁵
 - ❖ If pathogen is present --> amplification --> fluorophore hydrolysis --> [fluorescent signal](#)
 - ❖ Increasingly common due to reduction of potential contamination
- ❖ High [specificity](#) and [sensitivity](#)
- ❖ Can be done with [small sample volumes](#) of CSF²⁷

IMAGE SOURCE: PEXELS¹⁴ 2020. IMAGE FREE FOR USE

HAND HOLDING PETRI DISH



RESULTS

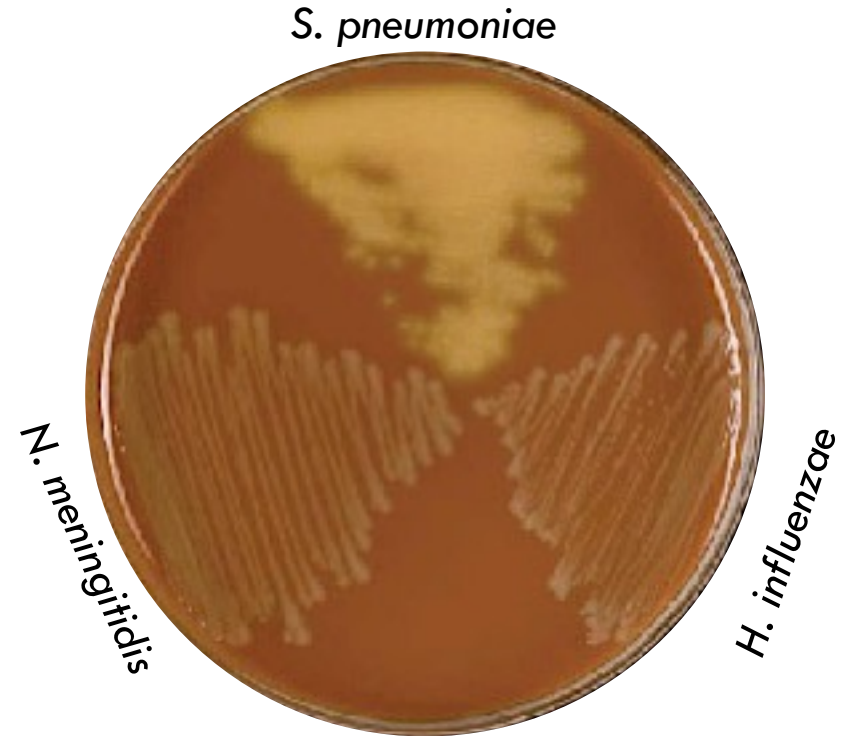
What are the results expected from these tests allowing for the identification of the bacteria named in this case?

CULTURE

Organism	Blood Agar Culture		Chocolate Agar Culture	
	Colony morphology	Hemolysis	Colony morphology	Hemolysis
<i>N. meningitidis</i> ^{30,33}	Round, smooth, moist, glistening, convex, grey	No hemolysis	Large, colourless or grey, opaque	No hemolysis
<i>H. influenzae</i> ^{33,36}	No growth without supplementation	No hemolysis	Large, colourless or grey, opaque	No hemolysis
<i>S. pneumoniae</i> ^{32,33}	Small, grey, moist, mucoidal	Alpha-hemolysis (green)	Small, grey, moist, mucoidal	Alpha-hemolysis (green)

Note: *N. meningitidis* and *H. influenzae* appear similar on chocolate agar plates. Perform NAD growth factor test to confirm *H. influenzae*

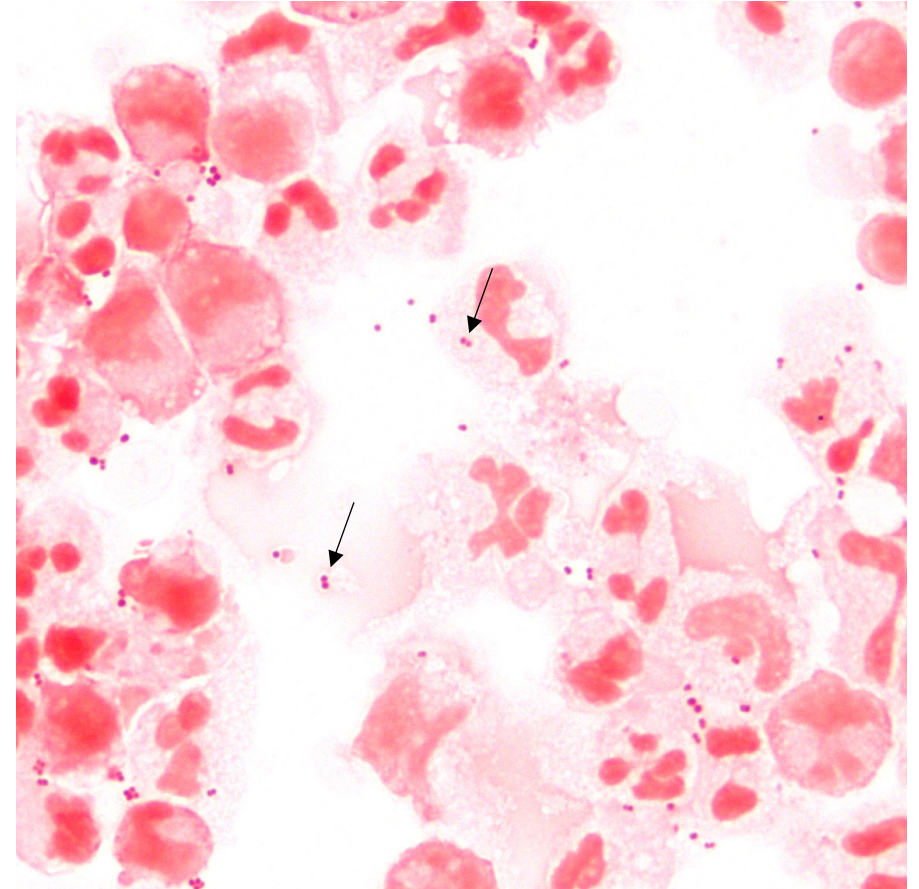
CULTURE



Note: *N. meningitidis* and *H. influenzae* appear similar on chocolate agar plates. Perform a NAD growth factor test to confirm *H. influenzae*.

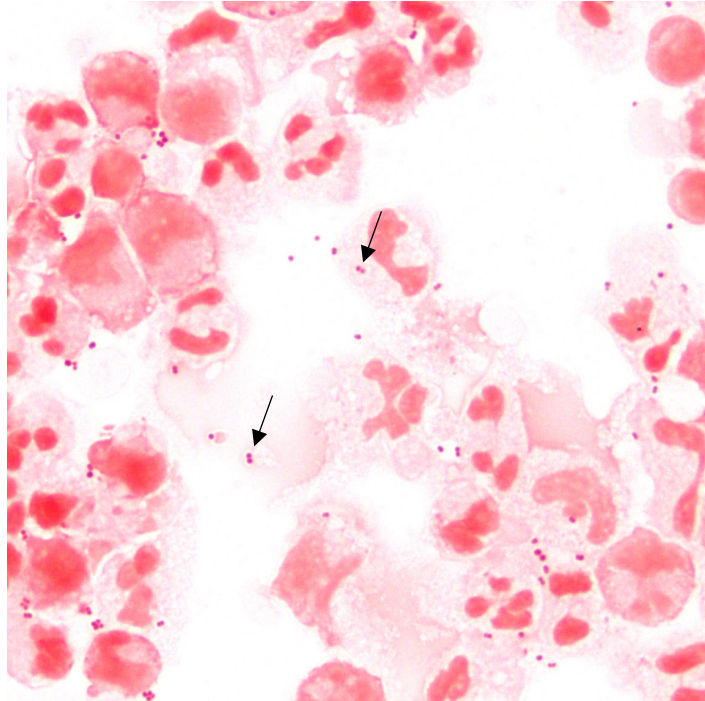
GRAM STAIN

- ❖ *N. meningitidis* is Gram-negative, should appear pink/red when Gram stained³⁰
 - ❖ Due to presence of outer membrane composed of lipooligosaccharides and lack of thick peptidoglycan layer³¹
- ❖ *H. influenzae* is also Gram-negative³⁶
- ❖ *S. pneumoniae* is Gram-positive³², should appear purple
 - ❖ Due to presence of thick peptidoglycan layer³¹



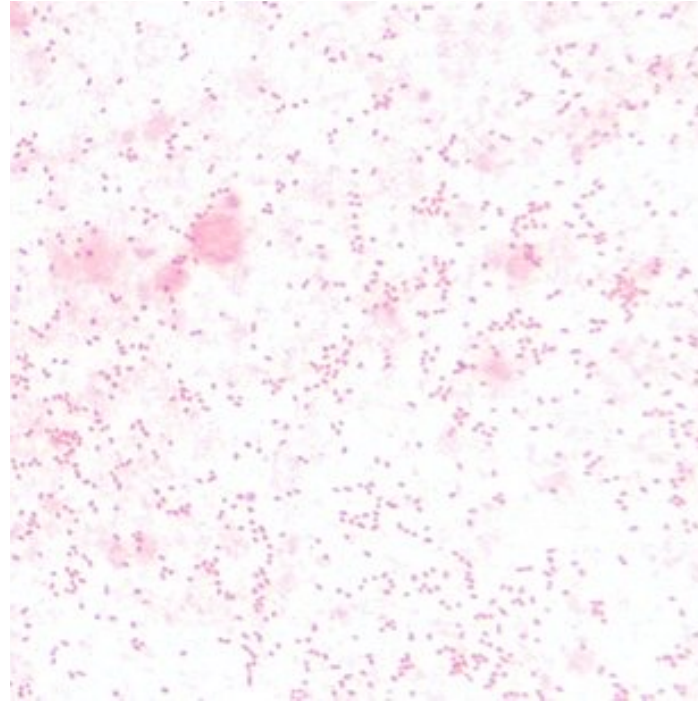
GRAM STAIN

Neisseria meningitidis



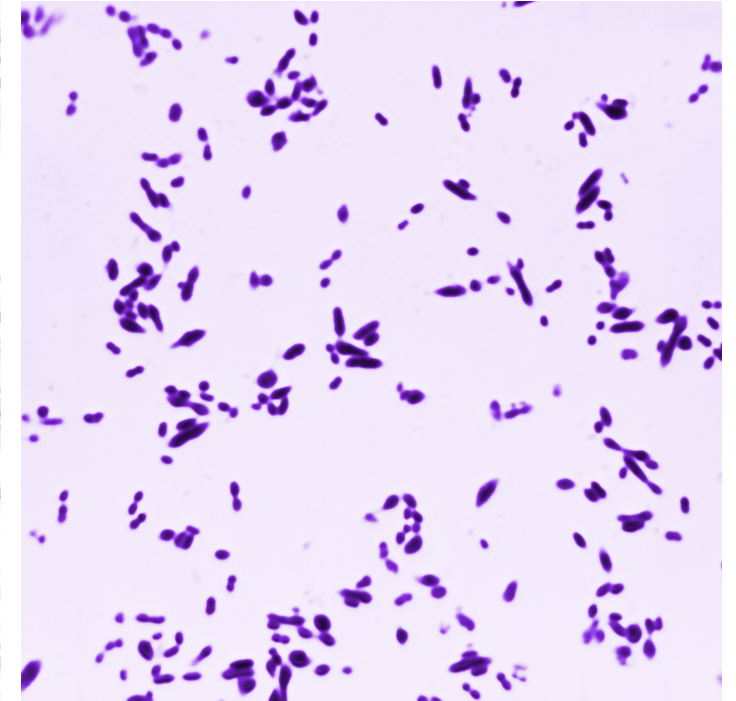
Gram-negative diplococci
Pair clustering³⁰

Haemophilus influenzae



Gram-negative, rod-shaped³⁶
Rounded ends (coccobacilli)

Streptococcus pneumoniae

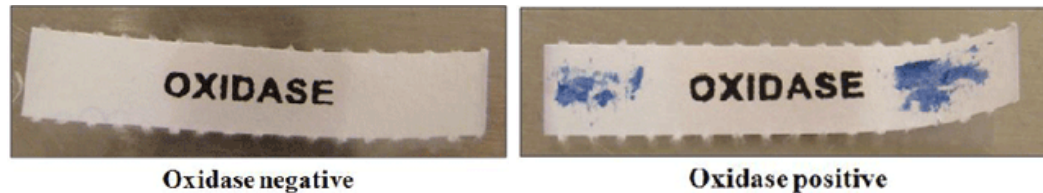


Gram-positive, lancet-shaped cocci³³
Pair clustering

BIOCHEMICAL TESTING

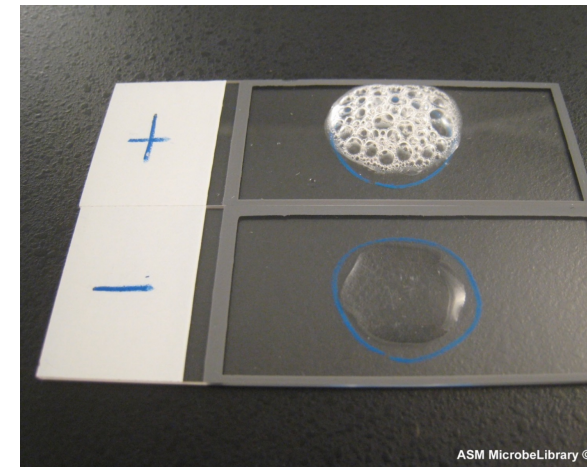
Kovac's Oxidase Test

- ❖ *N. meningitidis* and *H. influenzae* express cytochrome c oxidase --> positive oxidase test^{30,36}
 - ❖ Filter paper changes to blue
- ❖ *S. pneumoniae* does not express oxidase --> negative test³²
 - ❖ Filter paper remains colourless



Catalase Test

- ❖ *N. meningitidis*, *H. influenzae*, and *S. pneumoniae* do not express catalase³²
 - ❖ No bubbling observed when hydrogen peroxide added



CARBOHYDRATE UTILIZATION

❖ *N. meningitidis* ferments glucose and maltose
(colour change red --> yellow)³⁰

❖ Does not ferment lactose or sucrose

❖ *H. influenzae* ferments glucose and maltose³⁶

❖ Does not ferment lactose or sucrose

❖ *S. pneumoniae* ferments glucose, lactose, maltose, and sucrose³²



Maltose

Glucose

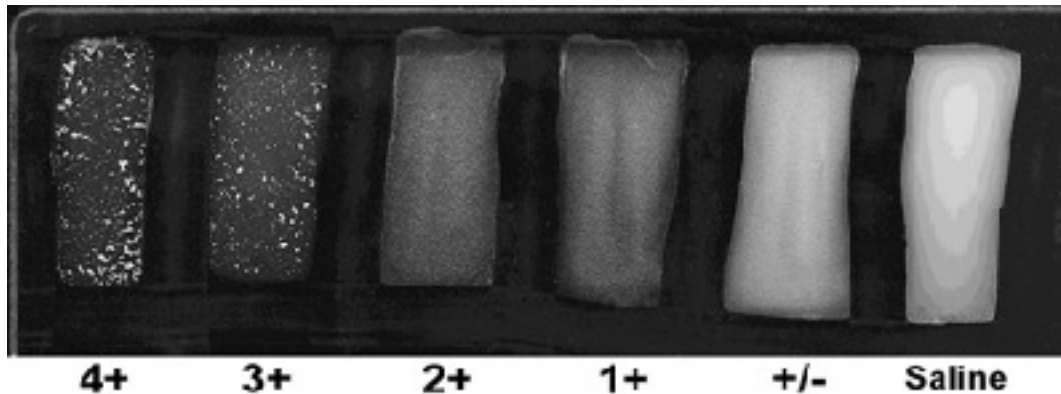
Lactose

Sucrose

AGGLUTINATION TESTING

Agglutination Serogrouping

- ❖ Used to identify **serotype** of *N. meningitidis*³⁰
- ❖ A rating of **3+** or **4+** indicates the presence of the unique serotype of *N. meningitidis*

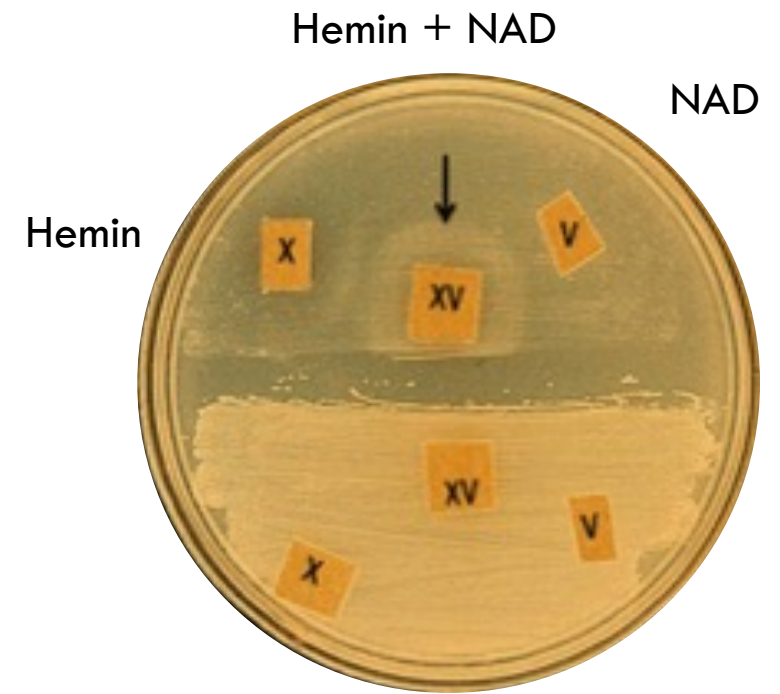


Latex Agglutination

- Positive test indicated by **clumping** seen within **5-10 seconds**
- Typically used as a **secondary method** of testing if³⁷:
 - Gram stain is **inconclusive**
 - Or meningococcal disease suspected
- Prone to **false negatives**³⁷
 - Positive in **60%** of *S. pneumoniae* cases
 - Positive in **93%** of *H. influenzae* cases
 - Positive in **39%** of *N. meningitidis* cases

HEMIN AND NAD GROWTH FACTOR TEST

- ❖ *N. meningitidis* and *S. pneumoniae* do not require hemin or NAD to grow
- ❖ *H. influenzae* requires hemin and nicotinamide-adenin-dinucleotide (NAD) to grow³⁹
 - ❖ No growth seen around disk containing only hemin
 - ❖ No growth around disk containing only NAD
 - ❖ Growth seen around disk containing hemin + NAD



TESTS TO CONFIRM *S. PNEUMONIAE*

Optochin test

- ❖ *N. meningitidis* and *H. influenzae* do not have their growth inhibited in the presence of optochin
- ❖ *S. pneumoniae* is optochin-sensitive³²
 - ❖ If zone of inhibition >14 mm, organism is considered sensitive
 - ❖ Pathogen is likely to be *S. pneumoniae*

Bile solubility test

- ❖ *N. meningitidis* and *H. influenzae* are not bile soluble
 - ❖ Negative test = solution remains turbid after 10 minutes, and even after 2 hours
- ❖ *S. pneumoniae* is bile soluble, unlike other alpha-hemolytic streptococci³²
 - ❖ Positive test = solution becomes clear/less turbid after 10 minutes

POLYMERASE CHAIN REACTION TEST

- ❖ Can be used to pinpoint presence of *N. meningitidis*, *H. influenzae*, and *S. pneumoniae* with **high specificity** and **sensitivity**³⁵
- ❖ Can also **detect serotype**
- ❖ *N. meningitidis* - expresses gene encoding the copper, zinc superoxide dismutase, *sodC*³⁵
- ❖ *H. influenzae* - expresses gene encoding the surface-exposed lipoprotein, protein D, *hpd*³⁵
 - ❖ Can be found in all *H. influenzae* serotypes
- ❖ *S. pneumoniae* - targets a specific segment of gene encoding autolysin, *lytA*³⁵

Target	Primer or probe name	5' to 3' nucleotide sequence
<i>N. meningitidis</i>		
<i>sodC</i>	F351	GCACACTTAGGTGATT TACCTGCAT
	R478	CCACCCGTGTGGATCATAATAGA
	Pb387	CATGATGGCACAGCAACAAA TCCTGTTT
<i>H. influenzae</i>		
<i>hpd</i>	hpdF822	GGTTAAATATGCCGATGGTGTTG
	hpdR952	TGCATCTTTACGCACGGTGTA
	Pb896i	TTGTGTACACTCCGT" T"GGT AAAAGAACTTGAC
<i>S. pneumoniae</i>		
<i>lytA</i>	F373	ACGCAATCTAGCAGATGAAGCA
	R424	TCGTGCGTTTTAATCCAGCT
	Pb400i	TGCCGAAAACGC" T"TGATAC

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