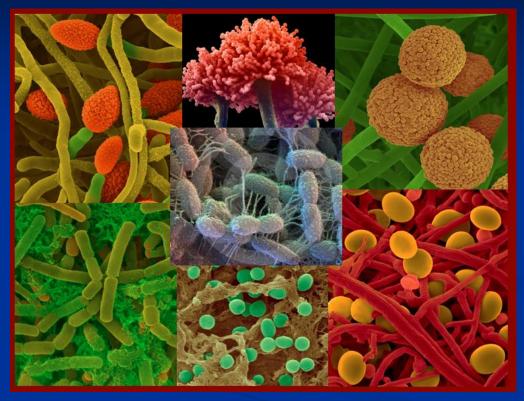
Biology Lab 2 Assessing fundamental symbiotic relationships between plants and soil microorganisms

Soil biodiversity

- Soil probably harbours most of our planet's undiscovered biodiversity (Tiedje et al., 1999)
- 1 tbsp. soil contains more microbes than there are people on earth
- "Microbial life can easily live without us; we, however, cannot survive without the global catalysis and environmental transformations it provides (Falkowski et al, 2008"



- Microorganisms can be free-living or in symbiosis
- Key symbiotic associations between plants & microbes
- N₂-fixing nodules (plants and bacteria)
- Mycorrhizae (plants and fungi)

Symbiotic bacteria Non-leguminous nodules

200 dicotyledonous plants form nonleguminous N₂ fixing nodules after infection with actinomycete -Frankia sp., all but 2 are woody and found in temperate regions and tropics Most prominent is Frankia sp. with Alder





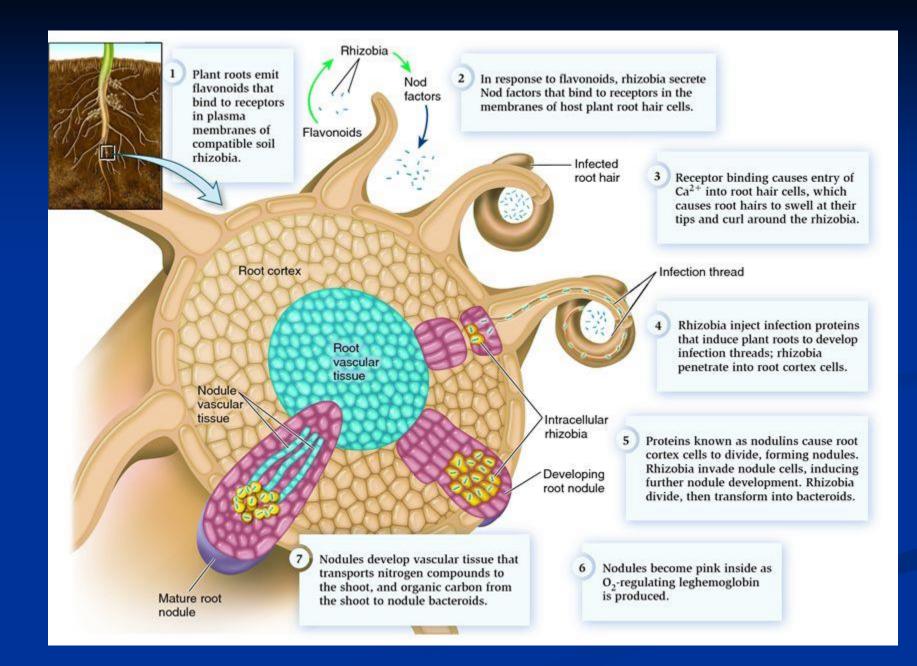


Symbiotic bacteria - Leguminous nodules

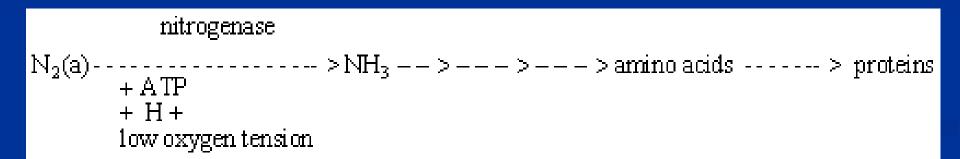
- Herbaceous legumes infection is through root hairs - host specific recognition signals recognised by Rhizobium
- > Peas
- Beans
- Clover
- > Vetch
- > Alfalfa
- > Soybean
- Lentils
- Chickpeas

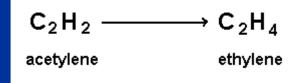






Biochemical mechanism of N_2 fixation

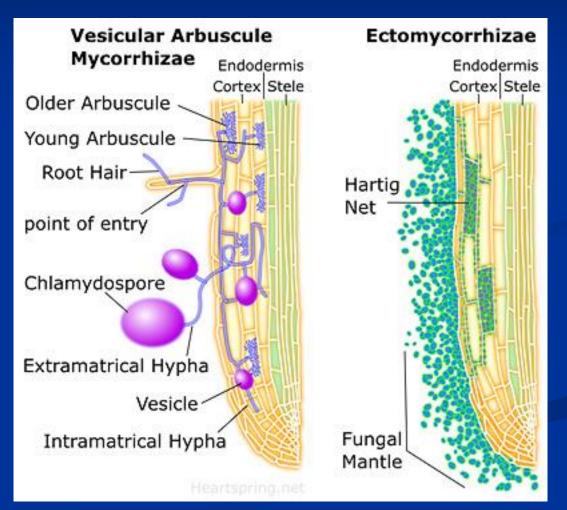




Nitrogenase also catalyses reduction of acetylene

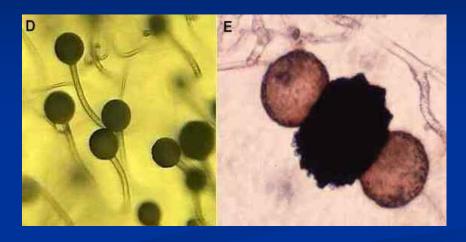
Symbiotic fungi Mycorrhizae

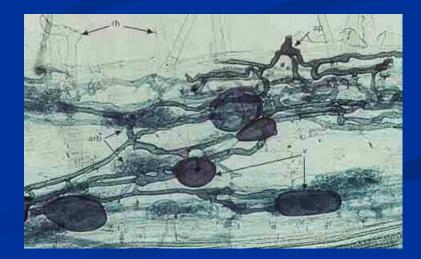
- Most plants are mycorrhizal: fungus receives C for growth, plant greater acquisition of nutrients
- Mycorrhizal fungi exist as spores/vegetative propagules in root fragments
- Germination & growth stimulated by root exudates chemotropism to root via exudates
- 3 types
 Arbuscular
 Ecto
 Ericoid



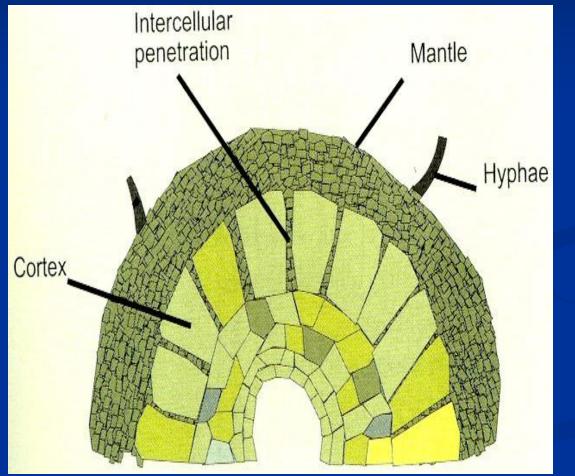
Symbiotic fungi - Arbuscular Mycorrhizae

- Arbuscular Mycorrhizae
- Found on vast majority of wild and crop plants, exceptions are Brassica. Some tree species eg cedar, maple, alder, majority tropical tree species
- Fungi belong to Zygomycete group
- Majority fungal mycelium is internal penetrates cortical cells
- Site of penetration is top right, fungus produces pre-infection swelling (appressorium, ap), grows between root cells and forms arbuscules (arb) and swollen vesicles (v).
- > Arbuscules sites of nutrient exchange
- > Vesicles used for storage.

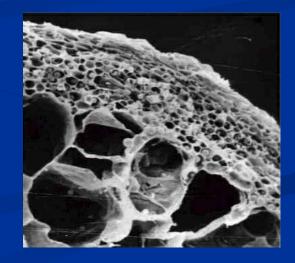




Symbiotic organisms Ecto Mycorrhizae







Symbiotic fungi – Ecto Mycorrhizae

- Predominate in trees particularly in Pine, Beech, Spruce, Fir and Birch
- Pine & oak are obligately mycorrhizal
- Fungi belong to Ascomycete and Basidiomycete groups – many mushroom producers
- Majority of fungus is external - forming hyphal mantle around root, external mycelium can extend many metres through soil and inter-connect trees











Outline

Skills

Isolation and estimation of % infection of plant roots with mycorrhizae, staining and microscopy techniques; morphotyping of ecto-mycorrhizae; visualization of root nodules and their viability; calculation of N₂ fixation rates

Objectives:

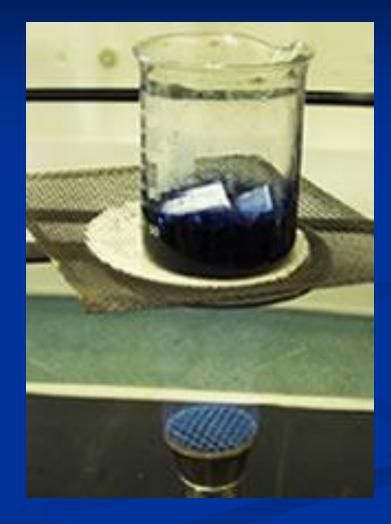
To observe, characterize and enumerate % colonization of plant roots by mycorrhizae using staining, microscopy techniques and morphotyping. To identify root nodules, enumerate and estimate N₂ fixation rates.

Methods

- Clear (boil in KOH) and stain herbaceous roots with ink and vinegar and estimate % AM infection using microscopy and the grid-line intersection method
- Wash fine roots of trees and observe ECM root tips using a dissecting microscope. Use colour atlas of mycorrhizae to morphotype ECM and estimate % infection of roots by different mycorrhizal fungi. Estimate ECM abundance and richness.
- Wash root nodules from legumes and trees. Calculate nodule mass.
 Determine % active (brown/red) vs inactive (white/green) nodules by dissection.
- Use known BNF rate per nodule to calculate BNF per plant per day

Clearing and staining of roots

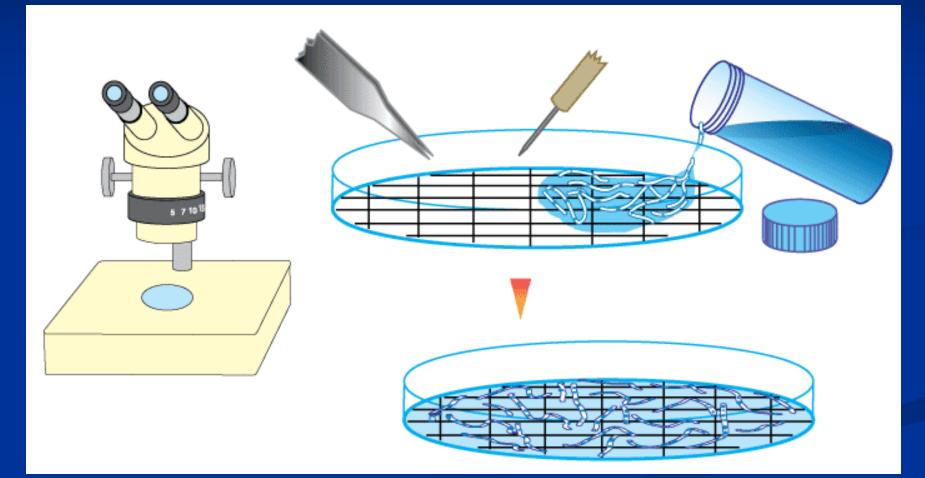




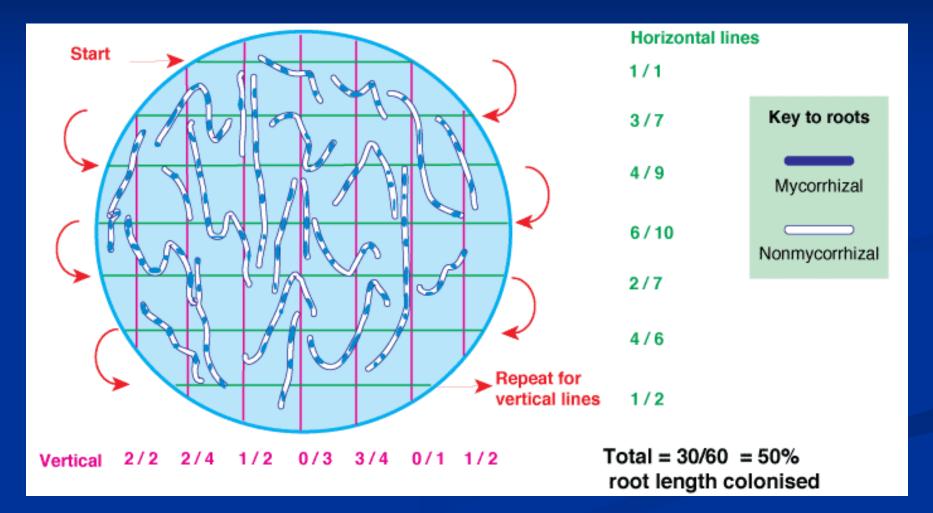
Clear with boiling KOH !

Stain with black ink and white vinegar

Put stained roots in petri dish with grid lines, randomly disperse & observe under dissecting microscope



Assess % mycorrhizal colonization using the gridline intersection method



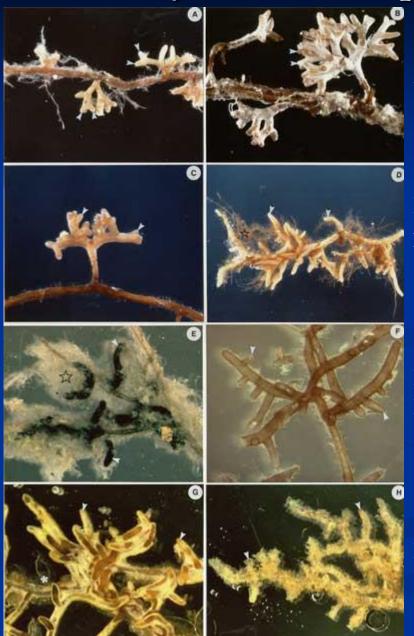
Examples of ectomycorrhizal morphotypes

Pine & Suillus brevipes

Pine & Boletus edulis

Eucalyptus & Tylopilus sp.

Eucalypt & Pisolithus sp.



Pine & Amanita muscaria

Eucalyptus & Astraeus pteridis

Eucalyptus & Redellomyces sp.

Eucalypt & Amanita sp.

Nodules of trees and legumes



Alder



Clover

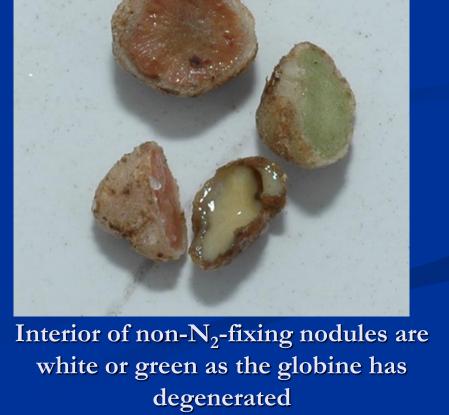
Nitrogenase is an O_2 sensitive enzyme. Low O_2 is realized through compartmentation in cyanobacteria, active respiration (in *Azotobacter*), synthesis of leghemoglobin (in *Rhizobium* legume).

Leghemoglobin is a macromolecule synthesized by both symbiotic partners - *Rhizobium* synthesizes the heme portion, and the plant the globine. Like human hemoglobin, leghemoglobin fixes O₂.

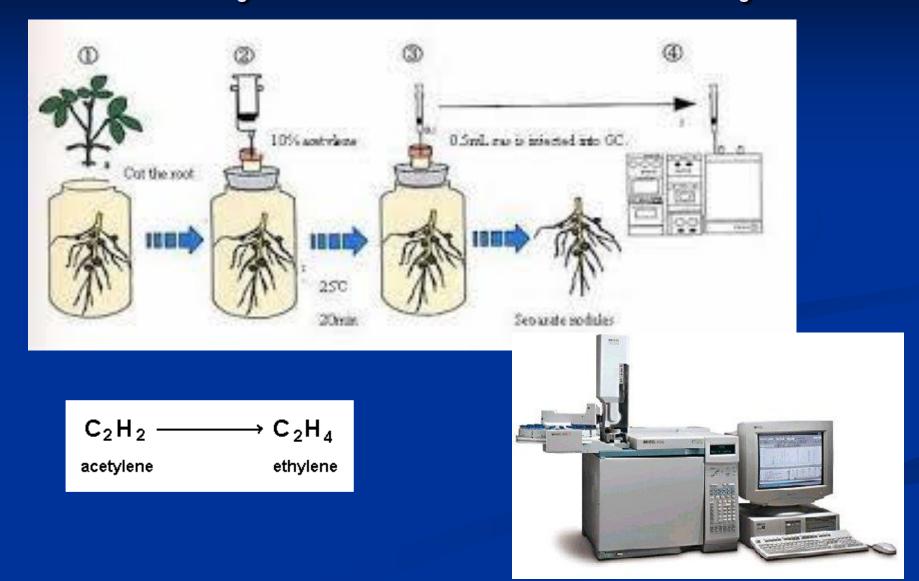
S Jennifer Dean

Red color of open nodule shows healthy rhizobia

Interior of active (i.e., N₂-fixing) nodules are red or brown



Acetylene reduction assay



You have been given data on N_2 fixation rate per g nodule (will see GC in week 3 in BEG labs)

Lab week 2

From week 1:

- Identify meso-fauna using microscopy and keys
- > Determine % feeding activity in fauna in your soil from bait strips
- Enter faunal species diversity into table on connect (so everyone can calculate E (evenness) of faunal communities) BY TUESDAY NOV 19
- Week 2
- > Stain and assess % AM infection of herbaceous roots
- Assess diversity of ECM morphotypes on woody roots and the % ECM infection of each type
- Count nodules on roots, dissect nodules to determine if nodules are active, weigh total nodule mass per plant
- Enter %AM infection of roots, %ECM infection of roots, ECM diversity into table on connect (so everyone can compare different plant/soils) BY FRIDAY NOV 22
- Enter nodule numbers, mass and % active into into table on connect (so everyone can compare different plant/soils) BY FRIDAY NOV 22

WEEK 1 lab submission due FRIDAY NOV 22 11.59PM and WEEK 2 lab submission due MONDAY NOV 25 1PM

Further reading

- Fundamentals of Soil Ecology, Second Edition (1994) by David C. Coleman, Jr., D. A. Crossley, Paul F. Hendrix
- Soil Microbiology, Ecology and Biochemistry, Third Edition (2006) by Eldor A. Paul
- Biological Diversity and Function in Soils (2006) by <u>Richard Bardgett</u>, <u>Michael Usher</u>, <u>David</u> <u>Hopkins</u>
- **The Biology of Soil: A Community and Ecosystem Approach (2005)** by <u>Richard D. Bardgett</u>
- Falkowski PG et al., 2008. The microbial engines that drive earth's biogeochemical cycles. Science 320, 1034-1039.
- Digital learning centre for microbial ecology http://commtechlab.msu.edu/sites/dlc-me/index.html
- Fun facts about fungi http://www.herbarium.usu.edu/fungi/funfacts/factindx.htm
- The microbial world <u>http://helios.bto.ed.ac.uk/bto/microbes/index.htm#Top</u>